Polyols that Accumulate in Renal Tissue Uncouple the Plasma Membrane Calcium Pump and Counteract the Inhibition by Urea and Guanidine Hydrochloride

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Sorbitol and mannitol, two stereoisomeric osmolytes, inhibit the ATP-dependent Ca\(^{2+}\) transport in inside-out vesicles derived from basolateral membranes from kidney proximal tubules. This inhibition (IC\(_{50}\) = 400 and 390 mm respectively) cannot be attributed to an increase in Ca\(^{2+}\) permeability, since the rate of EGTA-stimulated Ca\(^{2+}\) efflux from preloaded vesicles is not modified by these osmolytes. In the presence of 1 m sorbitol or mannitol, Ca\(^{2+}\) uptake is inhibited by 70 and 75%, respectively. Since the Ca\(^{2+}\)-stimulated ATPase activity is unaffected, sorbitol and mannitol uncouple the Ca\(^{2+}\) transport from the ATPase activity. The inhibition of Ca\(^{2+}\) transport by these osmolytes is reversible, since the inhibition disappears when the vesicles are preincubated with 1 m sorbitol or mannitol and then diluted 25-fold in reaction medium to measure Ca\(^{2+}\) accumulation. On the other hand, these osmolytes protect the (Ca\(^{2+}\)+Mg\(^{2+}\))ATPase from the inhibition of Ca\(^{2+}\) transport and ATPase activity by urea and guanidinium. These data suggest that the high concentrations of polyols that renal cells accumulate during antidiuresis, may regulate Ca\(^{2+}\) transport across the plasma membrane. In addition, polyols may protect the (Ca\(^{2+}\)+Mg\(^{2+}\))ATPase from the deleterious structural effects of urea, a compound that also accumulates during antidiuresis.

Introduction

Organisms and cellular systems that are required to adapt to stress conditions such as high temperature, desiccation, or high concentrations of urea respond by concentrating one of several organic solutes such as sugars, polyols, amino acids and methyamines (Yancey et al., 1982; Crowe and Crowe, 1984; Somero, 1986). This accumulation is associated with the effectiveness of these osmolytes in minimizing protein denaturation in the face of stress (Crowe and Crowe, 1984; Carpenter and Crowe, 1989; Sola-Penna and Meyer-Fernandes, 1994). In terms of function they have been classified either as compatible solutes, which have little effect on protein function, or as counteacting solutes, which offset the structural and functional effects that deleterious solutes have on cell proteins (Bowlus and Somero, 1979; Yancey et al., 1982). These studies indicate that one class of these naturally occurring compounds is able to confer stability on proteins in vivo without modifying their biological activity.

It has been demonstrated that some osmolytes protect against the effects of urea on the function of membrane-bound (Ca\(^{2+}\)+Mg\(^{2+}\)) ATPases from sarcoplasmic reticulum and renal plasma membranes, and also modulate independently the catalytic cycle of these Ca\(^{2+}\)-transporting ATPases (de Meis and Inesi, 1988; Vieyra et al., 1989, 1991; Chini et al., 1991, 1992). They increase the apparent affinity for Pi during the phosphorylation reaction of rabbit sarcoplasmic reticulum (Ca\(^{2+}\)+Mg\(^{2+}\)) ATPase (Jorge-García et al., 1988; de Meis and Inesi, 1988; Chini et al., 1991), and increase the rate of ATP <--- Pi exchange catalyzed by (Ca\(^{2+}\)+Mg\(^{2+}\)) ATPase from renal plasma membranes (Vieyra et al., 1989, 1991). It has been postulated that they modify the equilibrium between the different conformations of the enzyme (Chini et al., 1991), and that they may exert their influence through a decrease in water activity and in

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protein solvation (de Meis, 1989; Chini et al., 1991).

A recent report has shown that a decrease in protein solvation promoted by carbohydrates can modify the equilibria among allosteric conformations of hemoglobin (Colombo et al., 1992), an indication that the flexibility and dynamic nature of proteins are affected by protein-bound water molecules in a way that can be modified by interaction with biological solutes. We have demonstrated that trehalose uncouples the Ca\textsuperscript{2+} transport from ATP hydrolysis by the (Ca\textsuperscript{2+}+Mg\textsuperscript{2+})-ATPase of renal plasma membranes (Sola-Penna et al., 1994). This solute is accumulated in several organisms during some stress conditions, and it interacts strongly with water and with polar groups of proteins and phospholipids (Crowe and Crowe, 1984; Crowe et al., 1987).

Kidney is the mammalian organ that accumulates several osmolytes under specific conditions such as antidiuresis. The accumulation of very high concentrations of urea (0.4–0.9 m, Knepper and Rector, 1991) would be lethal to this tissue. However, almost in parallel with the increase in urea, counteracting compounds, such as methylamines, also accumulate in this tissue (Yancey and Burg, 1989). The parallel accumulation of this class of compounds has been associated with a protection against the deleterious effects of urea in renal tissue (Yancey and Burg, 1989). Another family of compounds that are accumulated in parallel with urea is that of polyols, especially sorbitol, mannitol and inositol (Bagnasco et al., 1986). It is of interest to note that these osmolytes are the most effective, among a series of polyhydric alcohols, in stabilizing oligomeric enzymes and in enhancing the re-assembly of denatured subunits in the presence of high urea concentrations (Shifrin and Parrott, 1975). The counteracting solutes represented by the methylamine and polyol families are found in large amounts in the renal medulla (Bagnasco et al., 1986), and they are also present in the renal cortex (Gullans et al., 1988).

The present report is aimed to investigate whether compounds of the polyol family that accumulate in renal cells in antidiuresis conditions can, like the methylamines, counteract the inhibitory effects of urea and guanidinium on the activity of the Ca\textsuperscript{2+} pump of the basolateral plasma membranes of kidney proximal tubule cells. Our results show that sorbitol and mannitol uncouple ATP hydrolysis and Ca\textsuperscript{2+} transport without increasing the Ca\textsuperscript{2+} permeability of the membrane, and that they protect the pump from inhibition by urea and guanidinium hydrochloride.

**Materials and Methods**

**Preparation of purified vesicles derived from basolateral membranes**

Basolateral membrane vesicles were isolated from sheep kidney proximal tubules by the Percoll gradient method (Grassl and Aronson, 1986). Compared with the initial homogenate this membrane fraction was enriched 9–12-fold in the specific activity of the basolateral membrane marker (Na\textsuperscript{+}+K\textsuperscript{+})-ATPase. Protein concentrations were determined using the Folin phenol reagent (Lowry et al., 1951) and bovine serum albumin as a standard.

\[ {^{45}}\text{Ca Uptake} \]

Exempt when otherwise noted the basic medium contained, in a final volume of 0.5 ml, 30 mM Tris (tris(hydroxymethyl)aminomethane)-HCl buffer (pH 8.5), 5 mM ATP, 5 mM MgCl\textsubscript{2}, 1 mM ouabain, 10 mM Na\textsubscript{2}SO\textsubscript{4}, 0.1 mM EGTA ([ethylenebis(oxyethylenenitrilo)tetraacetic acid] and 0.1 mM \[^{45}\text{Ca} \text{CaCl}_2\] (10 \textmu m free Ca\textsuperscript{2+}). The apparent \( K_{\text{Ca,EGTA}} \) used was 2.8 × 10\textsuperscript{7} (Fabiato and Fabiato, 1979). The experiments were carried out at 37 °C. Concentrations of osmolytes and other modifications in the medium are specified in the corresponding figure legends. \[^{45}\text{Ca} \text{Ca}^{2+}\text{ uptake} \] was started by the addition of membranes (protein concentration 0.2 mg/ml) and stopped by Millipore filtration (Martonosi and Feretos, 1964), using 0.45 \textmu m pore size filters. The \[^{45}\text{Ca} \text{Ca}^{2+}\text{ remaining in the vesicles} \] was counted in a liquid scintillation counter after the filters were washed twice with 10 ml of a cold solution containing 2 mM La(NO\textsubscript{3})\textsubscript{3}, 100 mM KCl, and 20 mM MOPS (4-morpholinepropanesulfonic acid)-Tris (pH 7.0).

\[ (\text{Ca}^{2+}+\text{Mg}^{2+})\text{ATPase activity} \]

ATPase activity was measured in the same conditions as Ca\textsuperscript{2+} uptake, except that the reaction
was quenched with acid, followed by adsorption of the non-hydrolyzed $\gamma$-$^{32}$P$\text{ATP}$ on charcoal (Grubmeyer and Penefsky, 1981). Following centrifugation at $4000 \times g$ for 30 min, an aliquot of the supernatant was withdrawn to measure the amount of $^{32}\text{Pi}$ released. Spontaneous hydrolysis of $\gamma$-$^{32}$P$\text{ATP}$ was measured in tubes run in parallel in which the enzyme was added after the acid. The $(\text{Ca}^{2+} + \text{Mg}^{2+})\text{ATPase}$ activity was the difference between the ATP hydrolysis measured in the presence of $\text{CaCl}_2$ and in its absence ($\text{EGTA}$ 1 mM). $^{32}\text{Pi}$ was counted in a liquid scintillation counter.

Results

Effects of sorbitol and mannitol on $\text{Ca}^{2+}$ uptake and on $(\text{Ca}^{2+} + \text{Mg}^{2+})\text{ATPase}$ activity catalyzed by inside-out vesicles derived from basolateral membranes of kidney proximal tubules

Fig. 1 shows that sorbitol (Fig. 1A) and mannitol (Fig. 1B) inhibit $\text{Ca}^{2+}$ uptake in a dose-dependent manner, with $I_{0.5} \approx 0.4$ m in both cases (empty circles). This effect is specific for the active $\text{Ca}^{2+}$ transport since the ATP-independent $\text{Ca}^{2+}$ binding is not modified by the addition of sorbitol or mannitol (data not shown). The same figures show (filled circles) that the ATP hydrolysis stimulated by micromolar $\text{Ca}^{2+}$ concentrations and measured under the same conditions as $\text{Ca}^{2+}$ transport is not inhibited by sorbitol (Fig. 1A) nor by mannitol (Fig. 1B). The uncoupling effect of sorbitol and mannitol on ATP-dependent $\text{Ca}^{2+}$ transport is completely reversible. The inhibition of $\text{Ca}^{2+}$ uptake disappears if
Effects of urea and guanidinium hydrochloride on 
Ca2+ uptake and on (Ca2+ + Mg2+)ATPase activity

Fig. 2 A shows that urea inhibits Ca2+ uptake in 
a dose-dependent manner, essentially in parallel 
with the inhibition of Ca2+-ATPase activity (empty 
and filled circles). The curve is biphasic, with a 
plateau between 0.2 and 0.6 M, a result that may 
be an indication that two different mechanisms are 
involved. Guanidinium hydrochloride, another 
agent considered to be a potent denaturant, in­
hibits both Ca2+ transport and ATP hydrolysis in 
a monophasic manner with an $I_{0.5} \approx 130$ mM 
(Fig. 2B). These results show that solutes of this 
type – in contrast to sorbitol and mannitol – do 
not uncouple Ca2+ transport from ATP hydrolysis.

In common with other counteracting solutes such 
as methylamines, polyols appear to act on specific 
domains of energy transducing enzymes (Sola-
Penna et al., 1994; Coelho-Sampaio et al., 1994), 
whereas urea and guanidinium hydrochloride pro­
mote a global inhibition of the pump. Their effects 
may be related to their ability to unfold the ATP-
pase molecule as a whole (Mashino and Fridovich, 
1987).

Effect of polyols, urea, and guanidinium on Ca2+ 
permeability

Inhibition of the ATP-dependent Ca2+ transport 
by sorbitol and mannitol might be associated with 
an increase in the Ca2+ permeability of the mem­
brane. However, it was found that a polyol con­
centration that promotes 50% inhibition of Ca2+ 
accumulation does not increase the rate constant 
for EGTA-induced Ca2+ efflux from vesicles pre-
loaded with Ca\textsuperscript{2+} (Fig. 3). The same figure shows a similar lack of effect of urea and guanidinium on the passive Ca\textsuperscript{2+} efflux. Since the effects of both classes of solutes on Ca\textsuperscript{2+} uptake can not be attributed to an increase in membrane permeability for Ca\textsuperscript{2+}, it may be concluded that they inhibit the unidirectional ATP-driven Ca\textsuperscript{2+} influx.

**Protection by polyols against inhibition of Ca\textsuperscript{2+} transport and ATP hydrolysis**

As shown in Fig. 2 A and 2 B, urea and guanidinium hydrochloride do not uncouple Ca\textsuperscript{2+} transport from ATP hydrolysis. Instead, and as mentioned above, they inhibit both reactions as a result of
their direct interaction with the structure of the 
(Ca^{2+} + Mg^{2+})\text{ATPase}. Fig. 4A and 4B, respectively, show that sorbitol and mannitol counteract 
the urea-induced inhibition of Ca^{2+} transport. Signif-
ificant protection is also conferred by these two 
polys in the guanidinium-induced inhibition 
(Fig. 5A and 5B). It should be mentioned that the 
Ca^{2+} transport by the vesicles is completely re-
stored when they are diluted 25-fold with urea-
free medium after incubation with 0.4 M urea but 
not after incubation with 1.5 M urea (inset to 
Fig. 4A and 4B, empty bars). In addition neither 
sorbitol nor mannitol modifies the complete resto-
ration of pumping activity of vesicles preincubated 
with 0.4 M urea, nor are they able to counteract 
the inhibition promoted by 1.5 M urea (inset to 
Fig. 4A and 4B, filled bars). With guanidinium 
chloride, which inhibits the enzyme with a simple 
monotonic concentration profile (Fig. 2B), both 
polys are able to preserve the functional state of 
the pump when they are present in the preincu-
bation medium even in the presence of a high con-
centration of the denaturating agent. Although 
Ca^{2+} uptake is inhibited when assayed after prein-
cubation of the vesicles in the presence of guanidi-
nium chloride with no sorbitol or mannitol (inset 
to Fig. 5A and 5B, empty bars), the pumping ac-
tivity attains the control value if the polys are 
also present in the preincubation media (inset to 
Fig. 5A and 5B, filled bars).

Finally, it can be seen that 0.4 M sorbitol 
(Fig. 6A and 7A) or mannitol (Fig. 6B and 7B), 
is as effective in counteracting also inhibitory ef-
fects of urea (Fig. 6) and guanidinium hydrochlo-
ride (Fig. 7) on the (Ca^{2+} + Mg^{2+})\text{ATPase} activity 
as they are for Ca^{2+} transport. Thus, although
these polyols uncouple Ca$^{2+}$ transport from ATP hydrolysis (Fig. 1), they are able to confer protection against the inhibitory effects of urea and guanidinium hydrochloride on the (Ca$^{2+}$ + Mg$^{2+}$)-ATPase.

**Discussion**

The protective role of organic solutes such as methylamines against protein damage by urea in biological systems (Yancey et al., 1982; Somero, 1986; Mashino and Fridovich, 1987; Jorge-Garcia et al., 1988; Vieyra et al., 1989, 1991) has been attributed to their ability to preserve the structural and functional integrity of proteins, probably because they promote compaction of the protein structure (Mashino and Fridovich, 1987). In this work, we show that polyols that are accumulated in renal cells at high concentrations under specific physiological conditions (Bagnasco et al., 1986) modulate the coupling ratio of the renal plasma membrane (Ca$^{2+}$ + Mg$^{2+}$)ATPase and confer protection against protein damage by urea and guanidinium hydrochloride.

In renal plasma membranes the uncoupling of ATP-dependent Ca$^{2+}$ transport and ATP hydrolysis (Fig. 1A and 1B) may reflect an interaction of sorbitol and mannitol with the enzyme phospholipid environment. Recently we have shown that trehalose also uncouples this transport system (Sola-Penna et al., 1994). This phenomenon may be related to the occurrence of strong interactions between polar groups of phospholipids and the hydroxyl groups of trehalose (Crowe and Crowe, 1984; Crowe et al., 1984; Sola-Penna et al., 1994). Since the chemical properties of the polyols used in this report are similar to those of trehalose, the sorbitol- and mannitol-induced uncoupling of ATP hydrolysis and Ca$^{2+}$ transport in renal plasma membranes may also be related to a decrease in membrane mobility promoted by interactions of the osmolytes with membrane phospholipids (Crowe and Crowe, 1988; Sola-Penna et al., 1994). Since the Ca$^{2+}$ binding site in different Ca$^{2+}$-ATPases is located in a hydrophobic region (Shull and Greeb, 1988; Clarke et al., 1989; Carafoli, 1991, 1992), interactions between these polyols and the phospholipids around the Ca$^{2+}$-binding domain might impair conformational changes of the enzyme associated with cation translocation. These results led to the conclusion that the long-range intramolecular linkage between Ca$^{2+}$-activated ATP hydrolysis and Ca$^{2+}$ transport is dissociated by the polyols.

The interaction of polyols with phospholipids around the ATPase may also increase the compaction of protein structure. Such an effect would confer protection against protein unfolding by urea and guanidinium hydrochloride (Robinson and Jencks, 1965), counteracting by this mechanism the inhibition of Ca$^{2+}$ transport and ATP hydrolysis (Fig. 4–7). Since thermodynamic studies have shown that mannitol at concentrations up to about 0.3 M has a structuring effect on water, while sorbitol has a structure-breaking effect (Stern and O’Connor, 1972), the uncoupling of Ca$^{2+}$ transport and ATP hydrolysis can not be explained by interaction of these compounds with water molecules.

The uncoupling promoted by sorbitol and mannitol (Fig. 1) suggests that in tissues where polyols occurs (Balaban and Knepper, 1983; Bagnasco et al., 1986; Balaban and Burg, 1987; Gullans et al., 1988), then may regulate physiological transport processes mediated by ATPases, including the plasma membrane (Ca$^{2+}$ + Mg$^{2+}$)ATPase responsible for the fine-tuned regulation of cytosolic Ca$^{2+}$ concentrations (Carafoli, 1991, 1992). During anti-diuresis, rabbits accumulate sorbitol and mannitol in the inner medulla (Bagnasco et al., 1986), where the concentration of urea can vary between 0.4 M and 0.9 M (Knepper and Rector, 1991), and there is evidence that Ca$^{2+}$ ions are involved in the process of intracellular osmoregulation (McCarty and O’Neil, 1992; Bagnasco et al., 1993). Since urea appears in the urine whereas polyols do not, it may be that the latter compounds play a significant role in both maintenance of intracellular osmotic balance (Bagnasco et al., 1986) and stabilization of ion-transporting ATPases in the presence of physiologically high urea concentrations.

Protection by sorbitol and mannitol against the effect of urea is almost complete in the first part of the inhibition curve (Fig. 4A and 4B), and appears to decrease above 1 M urea. Since the inhibition by 0.4 M urea is completely reversible whereas that promoted by 1.5 M is not, either in the absence or in the presence of one of the carbohydrates (inset to Fig. 4A and 4B), it may be concluded that counteracting effects of polyols occur...
only in the urea concentration range that is usually found in physiological conditions (Knepper and Rector, 1991).

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