Monophosphates and Cyclic Phosphates of Some Antiviral Acyclonucleosides: Synthesis, Conformation and Substrate/Inhibitor Properties in Some Enzyme Systems

Ryszard Stolarski\textsuperscript{a}, Jaroslaw M. Ciesla\textsuperscript{b}, and David Shugar\textsuperscript{a,b}

\textsuperscript{a} Department of Biophysics, Institute of Experimental Physics, University of Warsaw, 02-089 Warszawa, Poland
\textsuperscript{b} Institute of Biochemistry and Biophysics, Academy of Science, 02-532 Warszawa, Poland

Z. Naturforsch. \textbf{45c}, 293–299 (1990); received October 31, 1989

Acyclonucleosides, Monophosphates, Cyclic Phosphates, Conformation, Substrate/Inhibitor Properties

Chemical and enzymatic procedures are described for the synthesis of the monophosphates and cyclic phosphates of the antiviral acyclonucleoside 9-(1,3-dihydroxy-2-propoxymethyl)-guanine (DHPG), its 3-hydroxymethyl-4-hydroxybutyl analogue, the (R)- and (S)-epimers of 9-(3,4-dihydroxybutyl)guanine, and 9-(2,3-dihydroxypropyl)guanine.

The structures, and some conformational features, of all the foregoing, were determined by \textsuperscript{1}H and \textsuperscript{31}P NMR spectroscopy. Their substrate/inhibitor properties have been examined in several enzyme systems, including ribonucleases, snake venom phosphodiesterase, beef heart and higher plant cyclic nucleotide phosphodiesterases, nuclease P1, and 3'- and 5'-nucleotidases. The enzymatic results are considered in relation to the mechanism of the antiviral activity of the cyclic phosphate of DHPG.

Introduction

Since the discovery over 10 years ago of the antiviral activity of 9-(2-hydroxyethoxymethyl)-guanine (Acylovir, ACV), an acyclonucleoside analogue of guanosine [1], a multitude of purine and pyrimidine acyclonucleosides has been synthesized and tested for activity against various viruses (see e.g. [2, 3]). Interest in these unusual compounds has been further enhanced by the finding that some of them are potent inhibitors of enzymes involved in nucleic acid metabolism [4–7].

Whereas the antiviral activity of ACV, and some other acyclonucleosides is dependent on prior intracellular activation, viz. phosphorylation by viral-coded thymidine kinase (TK) in infected cells, there are others whose mode of action involves alternative pathways [8]. One of the most interesting is the cyclic phosphate of DHPG, 9-(1,3-dihydroxy-2-propoxymethyl)guanine (DHPG-cMP, see Scheme 1), which is a broad-spectrum \textit{in vitro} antiviral agent, the mechanism of action of which is not dependent on viral kinases and, indeed, is active against viruses which do not express such kinases [9].

The foregoing prompted us to prepare the monophosphates and cyclic phosphates of DHPG, C-DHPG, DHBG and HPG, the two latter of which may exist in two enantiometric forms, (R) and (S) (see Scheme 1), and to examine their conformational features, as well as their substrate/inhibitor properties in several relevant enzyme systems. This is an extension of previous investigations [10–12], based on the ability of the acyclic chains to mimic the conformation of some fragment of the pentose ring of the corresponding parent nucleoside, e.g. the acyclic chains of DHBG and HPG may be regarded as structural analogues of the “lower” portion of the 2'-deoxyribose ring. Such studies should contribute to a better understanding of the biological activities of these compounds, including their antiviral activities. It is also worth noting that the cyclic phosphates of DHPG, C-DHPG, DHBG and HPG might be considered as structural analogues of the signal molecule cGMP.

Abbreviations employed: ACV, 9-(2-hydroxyethoxymethyl)guanine; DHPG, 9-(1,3-dihydroxy-2-propoxymethyl)guanine; DHPG-cMP, DHPG-3',5'-cyclic phosphate; C-DHPG, carbo-DHPG, 9-(3-hydroxymethyl-4-hydroxybutyl)guanine; C-DHPG-cMP, C-DHPG-4',4'-cyclic phosphate; DHPAde, adenine analogue of DHPG; DHPAde-cMP, DHPAde-3':5'-cyclic phosphate; DHBG, 9-(3,4-dihydroxybutyl)guanine; DHBG-cMP, DHBG-3':5'-cyclic phosphate; DHBG-MP, DHBG-4'-monophosphate; HPG, 9-(2,3-dihydroxypropyl)guanine; HPG-cMP, HPG-2',3'-cyclic phosphate; PDase, phosphodiesterase; cPDase, cyclic nucleotide phosphodiesterase; RNase, ribonuclease.

Reprint requests to Jaroslaw A. Ciesla.

Verlag der Zeitschrift für Naturforschung, D-7400 Tübingen 0341–0382:90:0300–0293 $ 01.30/0
Scheme 1. Structures of investigated acyclonucleosides (left) and their cyclic phosphates (right). G = guanine; and a star indicates an asymmetric carbon: (A) DHPG and its 3':5'-cyclic phosphate; (B) C-DHPG and its 4':4''-cyclic phosphate; (C) (R)- and (S)-DHBG and their 3':4'-cyclic phosphates; (D) (S)-HPG and its 2':3'-cyclic phosphate.

Note: The numbering system for the acyclic chain carbons is in line with that for the carbons of the corresponding pentafuranose ring.

Materials and Methods

$^1$H NMR spectra of 0.02 M solutions were recorded on Bruker 270 and 400 MHz instruments at room temperature.

Thin-layer chromatography made use of Merck (Darmstadt, F.R.G.) F$_{254}$ plates with the solvent systems: (A) 1 M ammonium acetate:96% EtOH (2:5, v/v); (B) isopropanol:conc. NH$_4$OH:H$_2$O (7:1:2, v/v).

Sephadex DEAE A-25 was a product of Pharmacia (Uppsala, Sweden), Sephadex XAD-4 and Dowex 50W 1 × 8 were from Serva (Heidelberg, F.R.G.). POCl$_3$ was purified by simple distillation, trimethylphosphate by distillation under reduced pressure, and acetonitrile by successive distillation over P$_2$O$_5$ and CaH$_2$.

DHPG was a gift from Dr. J. G. Moffat. C-DHPG, (R)- and (S)-DHBG were gifts of Dr. N. G. Johansson, and (S)-HBG was a gift from Dr. A. Holy.

The cyclic phosphate of DHPAde, DHPAde-cMP, was prepared as previously described [12].

Beef heart cPDase (EC 3.1.4.17), snake venom PDase (EC 3.1.15.1), nuclease P1 (EC 3.1.30.1), RNases A (EC 3.1.27.5), T1 (EC 3.1.4.8) and T2 (EC 3.1.27.1), 3'-nucleotidase and 5'-nucleotidase were products from Sigma (St. Louis, U.S.A.). Potato tuber cPDase was a partially purified preparation, elsewhere described [13]. Values of $K_m$ and $V_{max}$ were obtained from Lineweaver-Burk plots with the aid of the Bisenthal-Cornish-Bowden algorithm, using an IBM PC program elsewhere reported [14].

(R)- and (S)-DHBG-MP. (R)-DHBG and (S)-DHBG were each phosphorylated with the aid of the wheat shoot nucleoside phosphotransferase system as elsewhere described [15]. The resulting 4'-monophosphates were isolated from the reaction mixture by chromatography on Whatman paper 3 MM with solvent B ($R_f$ of monophosphates 0.15, of DHBG 0.71). The products were eluted with water, and the eluates concentrated under vacuum and lyophilized (yields 40–50%).

(R)-DHBG-cMP. To a suspension of 480 mg (2 mmol) of (R)-DHBG in 10 ml triethylphosphate was added, portionwise and with constant stirring over a period of 5 h, 300 μl (3.3 mmol) POCl$_3$ [16]. The mixture was then poured onto ice and water, brought to neutrality with NaHCO$_3$, and loaded on a 3.3 × 30 cm column of DEAE-Sephadex A-25 (HCO$_3$). Elution was then conducted with a linear gradient of 0–0.8 M triethylammonium carbonate (2 × 21). Traces of non-reacted nucleoside were eluted with water, and the cyclic phosphate at 0.3–0.4 M, followed by ~5% of unidentified product. The cyclic phosphate was desalted, and freed from inorganic phosphate, on a Sephadex XAD-4 column by washing with water, and converted to the sodium salt on a Sephadex XAD-4 column by washing with water, and converted to the sodium salt on a column of Dowex 50 W 1 × 8 (Na$^+$). Precipitation with ethanol yielded 465 mg (75%) as a white powder, $R_f = 0.27$ on TLC, with solvent A ($R_f$ of (R)-DHBG = 0.52). The UV spectrum was identical with that of the starting product.

$^1$H NMR in (CD$_3$)$_2$SO with internal Me$_4$Si: δ (ppm) 10.64 (NH), 7.67 (H-8), 6.64 (NH$_2$), 4.05 (H-1'), 4.15 (H-3'), 4.02 and 3.56 (H-4'), 1.89...
(R') 31P NMR in 2H2O relative to external H3PO4: δ (ppm) 16.29. The product was resistant to alkaline phosphatase, but was hydrolyzed to the mononucleotide by potato tuber cPDase (see below).

(S)-DHBG-cMP. This was obtained by phosphorylation of (S)-DHBG (60 mg, 0.25 mmol) as for the (R)-epimer, above. In this instance, however, column chromatography led to a clear separation of the product from a minor unidentified product (∼2% yield). The cyclic phosphate was precipitated with ethanol to give 35 mg (45% of a white amorphous powder, chromatographically homogeneous, with solvent A, Rf = 0.27; UV, 31P NMR spectra identical with those of the (S)-epimer.

C-DHPG-cMP. Phosphorylation of 127 mg (0.5 mmol) of C-DHPG, as described above for (S)-DHBG, yielded 60 mg (35%) of a white amorphous powder, chromatographically homogeneous (Rf = 0.28; for C-DHPG, Rf = 0.58).

Alternative procedure, using the technique of Prisbe et al. [17]: to a solution of 127 mg (0.5 mmol) of C-DHPG in 100 ml acetonitrile was added 85 μl SnCl4 in 5 ml ethylene chloride. To the resulting clear solution was added portionwise, with stirring, pyrophosphoryl chloride in 50 ml acetonitrile, prepared by a procedure other than that of Prisbe et al. [17], viz. equimolar quantities (2.5 mmol) of POCl3 and water dissolved in 5 ml ethylene chloride. The reaction mixture was left for 24 h at room temperature, following which starting substance disappeared, with formation of one product. Chromatography on a column of DEAE-Sephadex A-25 was followed by desalting on a column of Sephadex XAD-4, and conversion to the Na+ salt, as above. Precipitation with ethanol gave 106 mg product (62% yield) in the form of white amorphous powder, chromatographically homogeneous, with a 1H NMR spectrum in (C2H3)2SO identical with that of the product described in the preceding paragraph: δ (ppm vs. internal Me4Si) 10.55 (NH), 7.70 (H-8), 6.48 (NH3), 3.95 (H-1'), 3.88 and 3.72 (H-4', H-4'' four protons), 1.63 (H-2'), 1.56 (H-3'). 31P NMR in 2H2O: δ (ppm vs. external H3PO4) = −2.68.

(D)-HBG-cMP. Phosphorylation of 56 mg (0.25 mmol) of HBG, as described above, yielded 42 mg (60%) of a white amorphous powder of the cyclic phosphate and 9 mg of an additional, unidentified product. The main product was chromatographically homogeneous, Rf = 0.31 with solvent A (Rf of (S)-HBG = 0.47). The UV spectrum was identical with that of the substrate. 1H NMR in 2H2O: δ (ppm vs. internal sodium 3-trimethylsilyl-(2,2,3,3-2H3)propionate) 7.88 (H-8), 4.37 and 4.31 (H-1'), ∼4.7 under H2O (H-2'), 4.41 and 4.07 (H-3').

Results and Discussion

Chemical phosphorylation

Phosphorylation of DHPG with POCl3 in triethylphosphate [16], as previously applied to phosphorylation of DHPAde [12], yielded as the major product the cyclic phosphate, DHPG-cMP (56%), and two minor products: 3',5'-bis-monophosphate and a mixture of 3'- and 5'-monophosphates. The structures were confirmed by 1H NMR spectroscopy [12]. The structure of the cyclic phosphate was additionally confirmed by the identity of its chromatographic properties and 1H NMR spectrum with those of the product of phosphorylation by the alternative procedure of Prisbe et al. [17], which leads to the cyclic phosphates of acyclonucleosides.

The foregoing results are at variance with those reported for phosphorylation of DHPG by Tolman et al. [9] and DHPAde by MacCoss and Tolman [18], the major products being the 3',5'-bis-monophosphates, accompanied by low yields of the 3':5'-cyclic phosphates. This discrepancy, previously alluded to [12], now appears due to differ-
ences in phosphorylating conditions. In our approach (see Experimental), POCl₃ was added portionwise to a solution of the acyclonucleoside in (CH₃O)₃PO, whereas in the procedure of Tolman et al. [9] and MacCoss and Tolman [18], the acyclonucleoside was added to a solution of POCl₃ in (CH₃CH₂O)₃PO (M. MacCoss, personal communication). This difference in phosphorylation conditions may therefore be profit from to prepare either the bis-monophosphates or the cyclic phosphates. Furthermore, under our experimental conditions, cyclic phosphate formation clearly must proceed via a monophosphate intermediate, followed by rapid nucleophilic attack at the phosphate moiety by the adjacent hydroxyl, i.e. under conditions where the POCl₃ concentration is low relative to the intermediate monophosphate.

Phosphorylation of C-DHPG with Yoshikawa method [16] yielded a single product, 4’:4”-cyclic phosphate (C-DHPG-cMP), the structure of which was confirmed by: (a) ¹H and ³¹P NMR spectroscopy in (C₂H₃)₂SO demonstrating the absence of OH groups and the characteristic coupling constants ~12 Hz between ³¹P and all the four protons at C(4’) and C(4”). (b) Resistance of the product to alkaline phosphatase; (c) identity of the chromatographic properties and ¹H NMR spectrum with those of the product of phosphorylation by the procedure of Prisbe et al. [17].

Further application of our phosphorylation method to the (R)- and (S)-enantiomers of DHBG, and the (S)-enantiomer of HPG again yielded, as the major products, the cyclic phosphates: (R)- and (S)-DHBG-3’:4’-cMP and (S)-HPG-2’:3’-cMP respectively, the latter previously reported by Holy [19].

It should be noted that, in contrast to the cyclic phosphates of DHPG and C-DHPG, the cyclic phosphates of the two DHBG enantiomers are 5-membered rings (linking a primary to a secondary hydroxyl) analogous to the 2’:3’-cyclic phosphates of nucleosides. The structures of the DHBG cyclic phosphates were established by: (a) ¹H and ³¹P NMR spectroscopy in (C₂H₃)₂SO, demonstrating the absence of OH groups, the presence of the phosphate group and the characteristic couplings between ³¹P and neighbouring protons; (b) resistance of both products to alkaline phosphatase; (c) hydrolysis of the cyclic phosphate rings by specific PDases to give the monophosphates (see below). In the case of DHBG and HPG, formation of the cyclic phosphates was accompanied by appearance of minor products (1%–10%) the identity of which is under study.

**Enzymatic phosphorylation**

With the aid of the wheat shoot nucleoside phosphotransferase system [15], the (R)- and (S)-enantiomers of DHBG were converted to the 4’-phosphates in 40–50% yield.

**Conformational aspects in solution**

In contrast to analogues with a 1,3-dihydroxy-2-propoxy(methyl) chain, for which there is a preference trans for substituents about C–O bonds [12], acyclic chains which may also mimic the “lower” portion of the pentose ring exhibit greater conformational flexibility (Scheme 2). For all the acyclonucleosides, and their cyclic phosphates, the values of the vicinal coupling constants ¹H–¹H (4.5 Hz–9.5 Hz) point to existence of a dynamic equilibrium of classical conformers with substituents gauche and trans about C–C bonds, and with comparable populations. This permits adoption by a given acyclic chain of conformations resembling those of the pentose ring.

The value of 11.8 Hz for the vicinal coupling constants ¹H–³¹P in C-DHPG-cMP shows that the cyclic phosphate ring exists as an equilibrium of two classical chair forms with identical populations. For one of these, C(2’) is gauche with respect to the cyclic phosphate ring oxygens, as prevails also for the O(4’) in DHPG-cMP and DHPAdcMP. In the second chair form, C(2’) is trans with respect to the ring oxygens as for the O(4’) in nucleoside 3’:5’-cyclic phosphates [12].

The conformation of the 5-membered phosphate ring of DHBG-cMP is in the envelope form with the phosphorus atom displaced from the plane of the remaining atoms, as deduced from the

![Scheme 2. Conformational equilibria about C–C and C–O bonds, indicated by arrows.](image-url)
coupling constants \(^{1}H-^{31}P\) in \(^{2}H_2O\), 5.8 Hz, 13.4 Hz and 8.1 Hz, respectively, for \(H(3')\) and the two \(H(4')\) protons. With HPG-cMP the \(^{1}H-^{31}P\) coupling constants point to an equilibrium of several forms similar to those of nucleoside 2':3'-cyclic phosphates [20]. More detailed conformational analyses are being undertaken.

**Enzymatic aspects**

None of the foregoing acyclonucleoside cyclic phosphates was a substrate for RNase A, T1 (which is specific for guanine residues) and T2, or for snake venom PDase. It should, however, be noted that the (S), but not the (R), enantiomer of the thymine analogue of HPG-cAMP, which can mimic the conformation of the corresponding \(\alpha\)-ribonucleoside 2':3'-cyclic phosphate, was earlier reported to be a substrate for several bacterial RNases [19]. The racemate of the foregoing was also claimed to be susceptible to RNase A [21], but the enormous concentrations of enzyme and substrate employed suggest that it is, at best, a very feeble substrate.

**Higher plant cPDase.** In line with previous observations that DHPAde-cMP, as well as 2',3'-seco-3':5'-cAMP, are substrates of potato tuber cPDase [11, 12], both enantiomers of DHBG-cMP, as well as (S)-HPG-cMP, were excellent substrates of this enzyme, each being converted to the monophosphate, readily dephosphorylated by alkaline phosphatase to the parent acyclonucleoside. From Lineweaver-Burk plots (see Fig. 1) the \(K_m\) values for both (R)- and (S)-DHBG-cMP were 0.5 mM, and for HPG-cMP 0.1 mM; while the \(V_{max}\) values were comparable to those for the natural substrates, 2':3'- and 3':5'-cAMP. By contrast, DHPAde-cMP was a poorer substrate (\(K_m\) ~10 mM and \(V_{max}\) about 10% that for DHBG-cMP); and DHPG-cMP was even less susceptible to hydrolysis, at a rate 2- to 3-fold lower than that for DHPAde-cMP.

Relevant to these findings are the known multi-substrate properties of potato (and other higher plant) cPDase, which hydrolyzes both 2':3'- and 3':5'-cyclic phosphates of nucleosides. We have elsewhere shown that mild alkaline treatment of the enzyme leads to selective loss of activity vs. 3':5'-cyclic phosphates, with full maintenance of activity vs. 2':3'-cyclic phosphates [13]. We now find that, following such selective inactivation of activity vs. nucleoside 3':5'-cyclic phosphates, there is no change in activity of the enzyme vs. both enantiomers of DHBG-cMP, and only a small decrease in the rate of hydrolysis of (S)-HPG-cMP. It follows that hydrolysis of these compounds is due solely to the activity of the enzyme vs. 2':3'-cyclic phosphates, consistent with the fact that they contain 5-membered cyclic phosphate rings. This was further conformed by the fact that the selectively inactivated enzyme did not hydrolyze DHPG-cMP or DHPAde-cMP, both with 6-membered cyclic phosphate rings.

**Beef heart cPDase.** Somewhat unexpected was the finding that both enantiomers of DHBG-cMP were converted to the mononucleotides by this mammalian enzyme, with \(K_m\) ~1 mM and \(V_{max}\) comparable to that for 3':5'-cAMP. DHBG-cMP was also slowly hydrolyzed by a crude extract of Ehrlich ascites cells (see also below). HPG-cMP was not a substrate of the beef heart enzyme.

**Nuclease P1:** This enzyme, at pH 5.3, hydro-
lyzed both enantiomers of DHBG-cMP to the mononucleotide. At pH 7.2 there was also formation of the nucleosides, consistent with the known 3'-phosphatase activity of this enzyme at the more alkaline pH [22]. (S)-HPG-cMP was hydrolyzed at about 50% the rate for DHBG-cMP.

*Other enzymes.* Both enantiomers of DHBG-4'-monophosphate were resistant to rye grass 3'-nucleotidase, but were very slowly converted to the nucleosides by venom 5'-nucleotidase (at about 1% the rate for 5'-AMP). The 4'-monophosphates also inhibited hydrolysis of DHBG-cMP by potato cPDase ($K_0 \sim 0.25$ mm).

The product of hydrolysis of DHBG-cMP by potato cPDase was resistant to venom 5'-nucleotidase, but was slowly converted to the nucleoside by 3'-nucleotidase. This suggests that the product of hydrolysis of DHBG-cMP by potato cPDase is the 3'-phosphate, for both the (R)- and (S)-epimers.

Quite surprising was the observation that the carba analogue of DHPG-cMP, *i.e.* C-DHPG-cMP, was resistant to all the foregoing enzymes, as well as to Ehrlich ascites cell extracts. This analogue also exhibited no inhibition of hydrolysis of 2':3'-cAMP or 3':5'-cAMP by potato cPDase.

*Antiviral activity of DHPG-cMP.* In contrast to the antitherpes activity of DHPG, DHPG-cMP exhibits potent activity not only against herpes, but also a number of other DNA viruses [23]. But, whereas the activity of DHPG is dependent on intracellular phosphorylation by viral thymidine kinase, this is not the case for DHPG-cMP [24]. As previously observed by us [11], as well as in the present study, DHPG-cMP is slowly converted to the monophosphate by cell extracts. But, as reported by Germershausen *et al.* [24], further phosphorylation to the triphosphate cannot account for the antiviral activity of this compound, and the mechanism of action of DHPG-cMP as an antiviral reagent remains to be clarified.

Furthermore, the nature of the enzyme(s) which hydrolyze DHPG-cMP in mammalian cell extracts has not been established. We have previously drawn attention to reports on the existence in mammalian cells of cyclic nucleotide phosphodiesterases with properties similar to those of our purified potato tuber cPDase [13], and consider that such non-conventional cPDase(s) may account for the slow hydrolysis of DHPG-cMP in mammalian cells. However, since such hydrolysis is still inadequate to account for the antiviral activity of this compound, other potential pathways must be considered. One possibility which has hitherto not been envisaged is that DHPG-cMP may be the active species as such, perhaps by acting as a signal molecule, like cGMP. It will be of interest to determine whether the cyclic phosphates of other antiviral acyclonucleosides, such as DHBG-cMP, also exhibit antiviral activity.

**Acknowledgements**

We are indebted to Dr. A. Holy for a sample of (S)-HPG, to Dr. Nils G. Johansson for samples of DHBG and C-DHPG, to Dr. J. Verheyden for a sample of DHPG, to Prof. Harri Lonnberg, Pentti Oksman and Prof. Heinrich Ruterjans for NMR facilities, and to the Alexander von Humboldt Foundation for support to one of us (R. S.) during the course of part of this research. This investigation was supported by the Polish Academy of Sciences (CPBR 3.13) and the Polish Cancer Research Program (CPBR 5.11).