Sequences Homologous to BAV Component of HL23V in Baboon Tissues

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HL23V-BAB, Sequence Homology, Competition Hybridization

The BAV component of HL23V virus (HL23V-BAB) hybridized to DNA from different Baboon tissues and appears to be an endogenous Baboon virus. Competition hybridization studies show that HL23V-BAB is closely related to viruses isolated from various Baboon species: P. hamadryas, P. papio, and P. cynocephalus. Competition hybridization studies also show that normal Baboon tissues contain more copies of proviral DNA than cells infected in vitro.

Introduction

Baboon endogenous viruses have been isolated from placenta, lung, kidney, and other tissues of several Baboon species. M-7 was the first baboon virus isolated from the placenta of Papio cynocephalus [1]. The second Baboon virus was isolated from the lymph node of a lymphomatous P. hamadryas. This virus is referred to as BILN. Many strains of Baboon endogenous virus have been isolated from lung, kidney and testicular tissues of Papio cynocephalus (BAB 8–1g, BAB 8K, BAB 455K), and from Papio anubis [3, 4]. The isolation of Baboon endogenous virus from the normal lung tissue of P. papio has also been reported. This virus is referred to as PP-1 Lu [5].

The virus HL23V isolated from the leukemic blood cells of a patient with acute myelogenous leukemia by Gallagher and Gallo [6] has been shown to be a mixture of Simian Sarcoma Associated Virus (SSAV) and Baboon endogenous Virus (BAV) [7, 8, 9, 10, 11]. The baboon component of HL23V (HL23V-BAB) was separated from SSAV and successfully propagated in human osteosarcoma cell lines [12]. Because HL23V-BAB was isolated from human leukemic cells and because it appears to be a new isolate of baboon virus, it was of interest to determine the sequence homology of HL23V-BAB in baboon tissues. In this paper, we show by competition molecular hybridization the homology that exists between HL23V-BAB and the endogenous virus from several Baboon species and that many copies of HL23V-BAB are present in Baboon tissues.

Materials and Methods

Tissues and cell lines

Liver and kidney tissues from P. hamadryas and spleen and brain tissues from P. papio were kindly provided by Dr. R.C. Gallo (National Cancer Institute, Bethesda, Maryland). AS/HOS/A7573 is a dog thymus cell line transfected with DNA from a human osteosarcoma cell line (AS/HOS) infected with HL23V-BAB. M-7 (8155) is a line of dog thymus cells producing M-7 Baboon virus. Both cell lines were kindly provided by Dr. P.D. Marksman, Litton Bionetics, Bethesda, Maryland.

Buffer

NTE is 0.1 m NaCl, 0.01 m Tris HCl and 0.001 m EDTA. NETS is 0.1 m NaCl, 0.01 m Tris, 0.1 m EDTA and 0.1 % SDS.

Preparation of 3H-labeled viral RNA

Confluent AS/HOS/A7573 cells were cultured in RPMI 1640 which contained 80 µCi/ml of [3H]-Uridine (ICN, Irvine, California), specific activity 28 Ci/mM, and 80 µCi/ml of [3H]-Cytidine (ICN, Irvine, California), specific activity 26 Ci/mM and 10% diazoyzed fetal calf serum. The cells were prelabeled for 4 h with radioactive medium which was discarded and then refed with radioactive medium which was collected at 6 h intervals for 18 h.

Viral RNA isolation

Virus was concentrated and partially purified from 16–20 liters of supernatant from AS/HOS/A7573 cells by centrifugation in a continuous flow
rotor (CF-32, Beckman). Concentrated virus was purified by equilibrium sedimentation centrifugation in a SW40 rotor (Beckman) through a 15–60% sucrose gradient in NTE, pH 7.4. The virus banded at a density of 1.16 g/ml. The viral RNA was extracted from purified virions with SDS and phenol, precipitated in presence of 20 μg of yeast tRNA with two volumes of ethanol and stored at −20°C. The viral RNA was pelleted, dissolved in 1.0 ml of NETS, pH 7.4 buffer, warmed at 37°C for 5 min and centrifugated on a gradient of 10–30% sucrose in NETS, pH 7.4 with 0.2 ml of 60% sucrose as cushion in a SW50.1 rotor (Beckman) at 40 K for 65 min at 20°C. C14-labeled 28 S rRNA was run in a parallel gradient. After sedimentation, the gradients were collected from the tube bottom. For unlabeled 60S RNA, OD at 260 was measured and fractions in the peak of 50–60S were pooled and precipitated with 2 volumes of ethanol and stored at −20°C.

For 3H-labeled RNA, 10 μl from each fraction were mixed with PCS scintillation fluid and counted in a Beckman scintillation counter. Fractions in the peak of 50–60S RNA were pooled and precipitated with 2 volumes of ethanol and stored at −20°C.

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Viral RNA

125I-labeled 60S HL23V-BAB RNA (specific activity: 108 cpm/μg) was kindly provided by Dr. R.G. Smith of the National Cancer Institute, Bethesda, Maryland.

Preparation of cellular DNA

The extraction and purification of cell DNA was as described by Baluda and Nayak [13].

Competition hybridization

The competition hybridization was performed as described [14–16]. The hybridization mixture contained 1 mg/250 μl of DNA, 800–900 cpm of 3H-labeled BAV 30S viral RNA (specific activity approximately 1×106 cpm/μg) and increasing concentrations (10–4 to 4 μg) of unlabeled 60S viral RNA or yeast RNA, 0.001 μm EDTA and 0.05% sodium dodecyl sulfate (SDS) in 0.4 M phosphate buffer (PB), pH 6.8. Mouse DNA was used to measure nonspecific binding of 3H viral RNA. The hybridization mixture was placed in tightly stoppered tubes, boiled at 100°C for 5 min and immediately transferred to a 65°C water bath. Hybridization was terminated after a Cot (mol of nucleotides × seconds × liter−1) of 40,000 was reached. At the end of hybridization, samples were processed as described [17].

Results

In sucrose velocity gradients RNA isolated from purified HL23V-BAB had sedimentation values of 50–60 S. After heating 50–60 S viral RNA at 80°C for 3 min, 30 S subunits were obtained. These results are in agreement with the reported sedimentation value of baboon virus [18].

Hybridization of 3H 30S HL23V-BAB RNA to cellular DNA from various baboon species and from infected cells

3H 30S HL23V-BAB RNA was hybridized with a large excess of DNA from normal Baboon tissue or from BAV-infected dog thymus cell lines. The ratio of cellular DNA to viral RNA was 106:1 on a mass basis and the ratio of proviral DNA to viral RNA was 10–15:1 assuming a single copy of proviral DNA per haploid cell genome.

The viral RNA was hybridized to a Cot of 40,000 mol sec−1. As shown in Table 1, 3H 30S HL23V-BAB RNA hybridized the most (64%) to DNA from P. hamadryas kidney or liver, 58% to DNA from P. cynocephalus lung, and an average of 52% to DNA from P. papio brain or spleen. There was 53% hybridization to DNA from AS/HOS/A7573 cells which produce HL23V-BAB and 51% hybridization to DNA from M-7 BAV.
Table I. Hybridization of \(^{3}H\) 30S HL23V-BAB RNA to cellular DNA from various baboon species and from infected dog cells.

<table>
<thead>
<tr>
<th>DNA Source</th>
<th>Percent Hybridization</th>
</tr>
</thead>
<tbody>
<tr>
<td>Royal baboon kidney ((P. hamadrys))</td>
<td>(64) (7)</td>
</tr>
<tr>
<td>Royal baboon liver ((P. hamadrys))</td>
<td>(63) ND</td>
</tr>
<tr>
<td>Yellow baboon lung ((P. cynocephalus))</td>
<td>(58) (4)</td>
</tr>
<tr>
<td>West african baboon brain ((P. papio))</td>
<td>(54) ND</td>
</tr>
<tr>
<td>West african baboon spleen ((P. papio))</td>
<td>(53) (13)</td>
</tr>
<tr>
<td>HL23V-BAB producing dog thymus cell line (\text{AS/HOS/A\text{7573}})</td>
<td>(54) (4)</td>
</tr>
<tr>
<td>M7 producing dog thymus cell line (\text{M7/8155})</td>
<td>(51) (9)</td>
</tr>
<tr>
<td>Uninfected dog thymus cell line</td>
<td>(10) (5)</td>
</tr>
<tr>
<td>Mouse embryos</td>
<td>(5) (5)</td>
</tr>
</tbody>
</table>

ND, not done.

Hybridization mixture contained 1 mg sonicated DNA, 800—1000 cpm of \(^{3}H\) 30S HL23V-BAB RNA (specific activity approximately \(1 \times 10^6\) cpm/\(\mu\)g), 0.001 M EDTA, 0.1% SDS in 125 or 250 \(\mu\)l of 0.4 M phosphate buffer, pH 6.8. Hybridization mixture was boiled for 5 min, transferred to a 65 \(^{\circ}\)C water bath and incubated to a Cot of \(4 \times 10^4\) mol sec \(^{-1}\). At the end of hybridization, samples were chilled in ice water and diluted with 2 \(\times\) SSC to a DNA concentration of 50 \(\mu\)g/ml and divided into two fractions. One-half was treated with 15 \(\mu\)g/ml of pancreatic RNase and 15 units/ml of T\(_1\) in 2 \(\times\) SSC at 37 \(^{\circ}\)C for 1 h and the other half served as control. Each sample was precipitated with 5% TCA, trapped on Whatman filters, washed, dried and radioactivity counted in a Beckman scintillation counter. The recovery of input radioactivity was 85—100%.

Kinetics of hybridization of \(^{3}H\) 30S HL23V-BAB RNA to DNA from \(P. hamadrys\) liver or \(\text{AS/HOS/A\text{7573}}\)

At a Cot of 40,000 mol sec \(^{-1}\), \(^{3}H\) 30S HL23V-BAB RNA hybridized 63% to DNA from the liver of a normal \(P. hamadrys\) (Fig. 1). The Cot\(_{1/2}\) of hybridization was approximately \(2 \times 10^9\) mol sec \(^{-1}\). The hybridization curve shows two components, 25% of the viral RNA hybridized by Cot of \(10^4\), and a second transition begins at Cot of \(10^3\). The Cot\(_{1/2}\) was determined for the entire heterogeneous reaction.

At a Cot of 40,000 mol sec \(^{-1}\), \(^{3}H\) 30S HL23V-BAB RNA hybridized 54% to DNA from \(\text{AS/HOS/A\text{7573}}\) cells (Fig. 1). The Cot\(_{1/2}\) of hybridization was approximately \(4 \times 10^3\) mol sec \(^{-1}\).

Kinetics of hybridization of \(^{125}I\) 60S HL23V-BAB RNA to DNA from \(P. papio\) or \(\text{AS/HOS/A\text{7573}}\) cells

At a Cot of 40,000 mol sec \(^{-1}\), \(^{125}I\) 60S HL23V-BAB RNA hybridized 53% to DNA from the spleen of a normal \(P. papio\) (Fig. 2). The Cot\(_{1/2}\) of hybridization was \(2 \times 10^9\) mol sec \(^{-1}\). At a Cot of 40,000 mol sec \(^{-1}\), \(^{125}I\) 60S HL23V-BAB RNA hybridized 55% to DNA from \(\text{AS/HOS/A\text{7573}}\) cells (Fig. 2). The Cot\(_{1/2}\) of hybridization was approximately \(4 \times 10^3\) mol sec \(^{-1}\).

Competition hybridization between \(^{3}H\) 30S HL23V-BAB RNA and homologous unlabeled 60S RNA with DNA from \(P. hamadrys\)

At a Cot of 40,000 mol sec \(^{-1}\) in presence of 1—4 \(\mu\)g of yeast RNA, \(^{3}H\) 30S HL23V-BAB RNA hybridized an average of 63% to DNA from a \(P. hamadrys\) liver and approximately 4% to normal murine DNA. Unlabeled 60S HL23V-BAB RNA at a concentration of 2—4 \(\mu\)g per 250 \(\mu\)l inhibited
the hybridization by 90%. The 50% inhibition level by homologous unlabeled 60S RNA was obtained at a concentration of \(5 \times 10^{-2}\) \(\mu g\) per 250 \(\mu l\). The normalized results are presented in Fig. 3.

**Competition hybridization between \(^3H\) or \(^{125}I\) 60S HL23V-BAB RNA and homologous unlabeled 60S RNA to DNA from West African Baboon (\(P.\ papio\))**

At a Cot of 40,000 mol sec\(^{-1}\), \(^{125}I\) 60S HL23V-BAB RNA hybridized an average of 42% to DNA from the spleen of a normal \(P.\ papio\) in presence of \(1 - 4\) \(\mu g/250\) \(\mu l\) of yeast RNA. Unlabeled 60S HL23V-BAB RNA at a concentration of \(5 \times 10^{-2}\) \(\mu g/250\) \(\mu l\) caused 50% inhibition. The normalized results are presented in Fig. 4. A competition hybridization experiment with brain DNA from \(P.\ papio\) and \(^3H\) 30S HL23V-BAB RNA gave identical results (data not shown).

**Competition hybridization between \(^3H\) 30S HL23V-BAB RNA and homologous unlabeled RNA with DNA from a cell line producing HL23V-BAB**

At a Cot of 40,000 mol sec\(^{-1}\), \(^3H\) 30S HL23V-BAB RNA hybridized an average of 54% to DNA from AS/HOS/A7573 cells in presence of \(1 - 4\) \(\mu g/250\) \(\mu l\) of yeast RNA. Unlabeled 60S BAV RNA at a concentration of \(2 - 4\) \(\mu g/250\) \(\mu l\) inhibited the hybridization by 98% and 50% competition was obtained at a concentration of \(1.5 \times 10^{-2}\) \(\mu g/250\) \(\mu l\). The normalized results are presented in Fig. 5.

**Competition hybridization between \(^3H\) 30S HL23V-BAB RNA and homologous unlabeled 60S RNA to DNA from dog thymus cells producing M-7 BAV**

At a Cot of 40,000 mol sec\(^{-1}\), \(^3H\) 30S HL23V-BAB RNA hybridized an average of 51% to DNA from M-7 (8155) cells in presence of \(1 - 2\) \(\mu g\) of yeast RNA. Unlabeled 60S BAV RNA from HL23V-BAB at a concentration of \(1 - 2\) \(\mu g/250\) \(\mu l\) inhibited the hybridization by 96%. Unlabeled 60S HL23V-BAB RNA at a concentration of \(1.6 \times 10^{-2}\) \(\mu g/250\) \(\mu l\) inhibited the hybridization by 50%. The normalized results are presented in Fig. 6.

**Discussion**

Competition hybridization between labeled and unlabeled RNA to template DNA in large excess is
used to determine the sequence homology between two species of RNA. In this study we have used DNA of different Baboon species to assess the sequence homology between HL23V-BAB and different Baboon proviruses. We have used 30S genomic RNA and large excess of cellular template DNA to determine the sequence homology of the entire viral genome. Competition hybridization results presented in this study have shown that HL23V-BAB is highly related to viruses of \(P. \) hamadrys, \(P. \) papio and M-7 (8155). Thus the baboon virus component of HL23 virus is an endogenous baboon virus widely distributed in different baboon species. Radioimmunoassay competition has shown that P-30 protein is highly conserved in \(P. \) cynocephalus, \(P. \) anubis, \(P. \) hamadrys \([4]\). The kinetics of hybridization of cDNA and mRNA to cellular DNA in excess can be used to determine the copy number of a gene from the Cot\(_{1/2}\) of the hybridization reaction \([20, 21]\). From the Cot\(_{1/2}\) of hybridization, it appears that DNA from normal baboon tissues contains approximately twice as much viral DNA as dog cells infected \textit{in vitro} which produce baboon virus.

The amount of unlabeled 60S BAV RNA required for fifty percent inhibition of hybridization of labeled viral RNA to different baboon DNAs reflects the concentration of BAV proviruses in the baboon tissues or infected cell lines. The concentration of unlabeled 60S BAV required for fifty percent competition of \(^3\)H HL23V-BAB 30S RNA was 3—4 times greater for baboon tissue DNA than for BAV infected dog thymus cell DNA (i.e. \(5 \times 10^{-2} \mu g/250 \mu l \) for baboon tissue DNA and \(1.5 \times 10^{-2} \mu g/250 \mu l \) for BAV infected cell line DNA). Therefore, the normal baboon tissues appear to contain 3—4 times more BAV proviral DNA sequences than the infected dog thymus cells, AS/HOS/A7573 or M-7 (8155) \([24]\). Our data are in agreement with the results of Benveniste and Todaro \([24]\) who have detected 5—15 copies of BAV proviral DNA per haploid cell genome in DNA from baboon tissues and 1 copy per haploid cell genome in DNA from BAV infected cell lines.

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